# Clinical Significance of Non-candidal Yeast like Genera with Special Reference to *Trichosporon* and *Malassezia*

**Microbiology Section** 

NAGESWARI RAJESH GANDHAM<sup>1</sup>, SAVITA VIVEKANAD JADHAV<sup>2</sup>, RABINDRA NATH MISRA<sup>3</sup>, CHANDA VIRENDRA VYAWAHARE<sup>4</sup>, NEETU SATISH GUPTA<sup>5</sup>

## ABSTRACT

**Introduction:** The incidence of opportunistic fungal infections is increasing world over. This is mainly due to increase in risk factors like prolonged antimicrobial use, prolonged catheterisations both urinary and vascular, increased patients with immune compromised conditions, co-morbidities like hypoalbuminemia or anaemia to name a few. Along with these newer aetiological agents are emerging as pathogens. Over the years, the predominance of *C. albicans* changed to non-albicans Candida and now Non-Candidal Yeast-like Genera (NCYG) are slowly emerging as pathogens. They include *Trichosporon, Malassezia* and others. Recently these are implicated in superficial, mucosal and systemic infections from immunocompetent patients also. Hence, this study was carried out to determine the incidence of NCYG in immunocompetent patients.

Aim: To isolate and identify NCYG in clinical samples.

Materials and Methods: This retrospective study included clinical specimen received over a one-year period from

immunocompetent patients from suspected fungal infections. All samples were processed for fungal follow-up as per standard protocol. This included microscopy, culture on Sabouraud Dextrose agar with and without antibiotics. Incubations were carried out at 37°C and at room temperature. Identification of growth was carried out as per follow-up for yeasts and automated VITEK ID was used for all isolates.

**Results:** Among various clinical specimens, fungal growth was obtained in 30.42% of samples. Of the yeast isolates, 85.47% were Candidal isolates. NCYG were isolated in 13.72%. Trichosporon was isolated in 46.15% of these NCYG. *Malassezia* was isolated in 26.93%. *Sporothrix, Pichia, Rhodotorula, Stephanoascus* and *Prototheca* were the other genera isolated.

**Conclusion:** The study highlights the presence of NCYG in clinical samples from immunocompetent patients. Identification and speciation of these emerging pathogenic yeasts should be done for better therapeutic management.

Keywords: Antimicrobial, Candidal isolates, Hypoalbuminemia, Immunocompetent, Infections

## **INTRODUCTION**

There has been an increase in incidence of infections caused by fungal aetiology. Mycotic infections may aggravate or add to the underlying disease process or by themselves be responsible for major distress, disability, morbidity or life-threating situations. For many years only a few genera of fungi were known to cause human infections. However, an increase in size of population at risk, along with the ubiquitous presence of fungi has lead to a great expansion in the fungal aetiology [1]. Genera and species once known to be commensals are increasingly implicated in disease. The emergence of these opportunistic infections has created a challenge in diagnosis and management of fungal infections and emphasised the importance of medical mycology. The protocols for culture and identification of which require dedicated effort and high index of suspicion on the part of the clinical microbiologist.

Mycotic agents of disease could morphologically be moulds or yeasts. Amongst the yeasts, the genera could be true yeasts or yeast-like and some genera resembling yeasts. Yeast-like genera are as designated as they are not true yeasts and morphologically show yeast and pseudo-hyphal forms. Of these yeast-like genera, the genus *Candida* is the most common opportunistic pathogen, research on which is quite comprehensive and ongoing. The other yeast and yeast-like genera include *Aureobasidium, Geotrichum, Malassezia, Pichia, Prototheca, Sporothrix* and *Tricosporon* to name a few [2,3]. Among these *Trichosporon spp*. could be misreported as non-albicans *Candida* [4]. The isolation and identification of these NCYG is not much reported. They need more tests and identification is tedious; however, the clinical significance of these is ever-increasing and they are the emerging opportunistic pathogens. This study was

planned with the aim to find the prevalence of these NCYG in clinical samples; To isolate and identify and speciate these genera.

## MATERIALS AND METHODS

This was a cross-sectional study carried out in the department of Microbiology of a tertiary care hospital in western Maharashtra. Samples received for mycology cultures from various clinical samples over a period of one year i.e., From January 2014 to December 2014; were included in the study.

Clinical suspected infections were classified as Mucocutaneous Infections (MCC), Urinary Tract Infections (UTI), Deep Seated Infections (DSI) and Blood Stream Infections (BSI). Samples for Mucocutaneous infections included skin, nail, vaginal secretions, ear discharge and pus from wound. Urine samples for UTI. DSI mainly included sputum, pleural fluid and broncho-alveolar lavages. BSI samples included blood and Central Venous Pressure (CVP) tips.

Samples were processed as per standard protocol for fungal follow-up. This included Microscopy, Culture, phenotypic manual identification tests and automated identification system VITEK2 Compact (Automated ID/AST by bioMerieux). Microscopy included wet mount, KOH mount, negative staining and Gram staining. All samples were cultured on four slants of Sabouraud Dextrose Agar (SDA); two with antibiotics and two without antibiotics. In addition, for mucocutaneous sample cultures were set up on SDA with olive oil at room temperatures for lipophilic fungi. One of each slant was incubated at 37°C and the other set at room temperature. Weekly readings were taken for four weeks. Positive cultures were identified by wet mount, gram stain, germ-tube test, CHROM agar urease test and VITEK 2 Compact system using YSTID cards [2,3].

# RESULTS

A total of 1982 samples from various clinical departments were included in the study. Amongst these fungal growths was obtained in 603 (30.42%) samples while the rest 1379 (69.58%) were negative for fungal growth [Table/Fig-1].

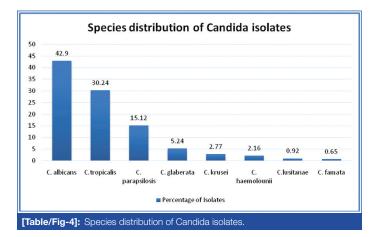
Total samples included	1982	
Culture positive for fungal pathogens	603 (30.42%)	
Culture negative for fungal pathogens	1379 (69.58%)	
[Table/Fig-1]: Details of samples and culture results.		

Among 603, 224 (37.15%) were identified as moulds. Yeast and yeast-like were identified in 379 (62.85%) isolates [Table/Fig-2].

Culture positive samples	Moulds/filamentous isolates	Yeasts and yeast-like isolates
603	224 (37.15%)	379 (62.85%)
[Table/Fig-2]: Details of isolates among Culture positive samples.		

**Yeast and yeast-like isolates:** Among the 379 isolates, 324 (85.47%) were identified as Candida isolates, three as true yeasts (0.79% all *Cryptococcus* sp.). The remaining 52 (13.74%) were identified as NCYG i.e., yeast and yeast-like genera other than *Candida* [Table/Fig-3]. Further, among Candida species, 139 (42.90%) were *C.albicans* and 185 (57.10%) were non-albicans Candida. The non-albicans Candida was speciated as *C.tropicalis*-98, *C.parapsilosis*-49, *C.glabrata*-17, *C.krusei*-9, *C.haemolounii*-7, *C.lusitanae*-3, and *C.famata*-2 [Table/Fig-4].

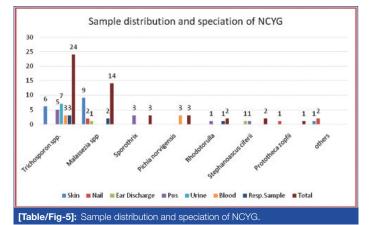
Total yeast and yeast-like isolates	379	
True yeasts	03 (0.79%)	
Candida isolates	324 (85.47%)	
NCYG	52 (13.74%)	
[Table/Fig-3]: Details of yeast and yeast-like isolates.		

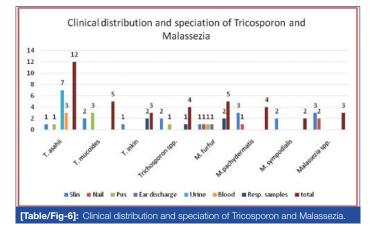


Thus 8.62% (52/603) of total culture positive were NCYG. Among yeast-like isolates the isolation rate was 13.72% (52/379). Among these isolates, 24 (46.15%) belonged to the genus *Trichosporon*. The next was *Malassezia* 14 (26.92%). The remaining were Sporothrix three, *Pichia norvigensis* three, two each *Rhodotorula* and *Stephanoascusciferii*, one of *Protothecazopfii* and others three.

Further speciation of *Tricosporon* showed *Trichosporonasahi* 12, *Tricosporonmucoides* five, *Trichosporoninkin* three, and others four. Of these six were from skin samples, seven from urine, five from pus and soft tissue samples, three from blood and three from respiratory samples [Table/Fig-5].

*Malassezia* speciation revealed *M. Furfur* five, *M.pachydermatis* four, *M.sympodialis* two and others three. Of these nine were from skin samples, two from nail, two from respiratory samples and 1 from ear discharge. The sample wise distribution of isolates is shown in [Table/Fig-6].





The male to female distribution of NCYG was 32:21 i.e., 1.5:1. Clinically isolates were from 33 MCC. This included 16 skin, five nail, 10 pus and soft tissue and two ear discharge. Among the rest, seven were from UTI, six were from BSI and 6 were from DSI. The DSI were pleural fluid, broncho-alveolar lavage and sputum.

#### DISCUSSION

In recent years infections caused by NCYG among the immunocompetent patients are being reported. High mortality rate has been reported from leukaemia patients. Among the yeast-like fungal pathogens Candida are the most predominant. They accounted for 85.47% of these isolates in the present study. The disease spectrum, pathogenicity, diagnostic methods and speciation of which have been well-researched and published. Although Non-albicans Candida are the most common fungal yeast-like pathogens encountered in clinical specimen, infections due to *Trichosporonspp.* are increasing [4,5]. Isolation of genera other than Candida requires more laboratory tests and time for their identification. The occurrence of these other genera in the present study was 13.74%. Therefore, they should not be ignored.

Sporothrix schenckii was isolated in three cases of skin and subcutaneous infection from hand. This a common site of isolation [6]. *Pichia* is a telomorph. The anamorphs of some *Pichia* species are *Candida* species *Candida inconspicua* and *Candida norvigensis* belong to fluconazole-resistant emerging species that are more frequently isolated from invasive infections. In addition to this increasing concern, these two species are difficult to identify and differentiate by routine conventional methods [7]. There were three isolates of *Pichia norvigensis*, all isolated from blood sample and identified by VITEK 2C system in the present study.

Previously considered non-pathogenic, *Rhodotorula* spp. have emerged as opportunistic pathogens that have the ability to colonise and infect susceptible patients. They are known to cause nail and skin infections and also fungemia in Central venous catheterised patients [8]. Two *Rhodotourula glutinis* isolated in this study were 1 each from respiratory and wound sample [Table/Fig-5].

In previous studies *Stephanoascus Ciferrii* has been isolated previously from blood, wound swab, pus, and aural discharge and nail sample [9]. In this study there were two isolates of *Stephanoascus Ciferrii* of which one isolate was from pus and other one was from ear discharge from immunocompetent patients.

*Trichosporonasahii* is an emerging fungal pathogen seen particularly in immunologically compromised patients requiring antifungal susceptibility testing to start effective treatment. In the present study clinical features of disseminated infections includes; septic shock, renal failure and cutaneous lesion.

The commonest genus in this study among NCYG was Trichosporon. It was 46.15% (24/52). Trichosporon belongs to class Basidiomycetes with unique morphological characters of budding yeast cell and true mycelium that disarticulate to form arthroconidia [2,4]. Previously only one species T.beiglii was reported as pathogenic. Now among the major human pathogens are-T. asahi, T.asteroides, T.mucoides, T.inkin, T.ovoides and T.Cutanei [10]. They are inhabitants of soil and can be a part of normal skin flora. They are responsible for superficial skin infections including white piedra. They may be responsible for systemic infections in patients with haematological malignancies or patients on steroids. These species are increasingly emerging as opportunistic pathogens mainly in immunosuppressed patients. Trichosporon may also be confused with Geotrichumsp as both produce arthroconidia [4]. They can be differentiated on basis of urease test and corn meal test. Further the isolates in present study were tested with VITEK ID system. By this T.asahii, T.mucoides and T.inkin were identified [Table/Fig-6].

T.asahii was the most common Trichosporon spp. isolated in 50%, 12 out of 24 isolates. This has been reported in other studies from India, Japan, Turkey and Brazil [4,11-13]. T.mucoides was second most common, 20.83%, five out of 24. T.inkin was isolated in 12.50%, three out of 24. Others were 16.67%, four out of 24. Trichosporon spp. are associated with blood stream infections and Nosocomial UTI [14,15]. In the present study Trichosporon was isolated from wide range of clinical specimen. Urinary tract infection was the most common, followed my mucocutaneous infections. Like our study Trichosporon spp. have been isolated from pus, soft tissue and respiratory specimen [4,16] [Table/ Fig-6] Though opportunistic they have also been reported from immunocompetent too. Risk factors such as prolonged multiple antimicrobials, indwelling catheters and co-morbidities such as anaemia and hypoalbuminemia may be contributory [17]. In the present study six isolates were from skin and nail samples, seven from urine, five from pus and soft tissue, three from blood and three from respiratory samples.

**Susceptibility testing for** *Trichosporon***:** There are no standard guidelines for interpretative criteria for minimum inhibitory concentration testing. Some workers have used the criteria for Candida species for reference. These findings suggest poor susceptibility to Amphotericin B while azoles have shown good potency against *T.asahii* isolates. Though emergence of resistance has been reported from different parts of the world, it has not been reported from India [4,15].

*Malassezia* is a lipid dependent yeast-like fungus also belonging to Class basideomycetes. The genus has at least seven species *M.furfur*, *M.globosa*, M.sympodialis, *M. obtuse*, *M.pachydermatis*, *M. slooffiae* to name a few. Direct microscopy of specimen exhibits the characteristic appearance of "spaggetti and Meatballs" [18]. Speciation was done on gross morphology, microscopy, urease test, lipophilicity and by automated system VITEK using YST ID cards. A total of 14 isolates were obtained. Male preponderance was seen similar to other studies [18,19]. *M.furfur* was the commonest five isolates, *M.pachydermatis* was second, four isolates, *M.sympodialis* two and others three. The isolates were most commonly from skin specimen followed by nail, respiratory and ear discharge. *Malassezia* can now be added to a growing list of normal skin flora organisms of low virulence that may cause mild recurrent skin infections and systemic infection in susceptible host [19,20]. Thus, *Trichosporon* and *Malassezia* accounts for 73.08% (38 out of 52) of the NCYG in this study [Table/Fig-5,6].

In previous studies, *Stephanoascus ciferii* has been isolated previously from blood. Wound swab, pus, and aural discharge and nail sample [9]. In this study there were two isolates of *Stephanoascus ciferii* of which 1 isolate were from pus and other one was from ear discharge from immunocompetent patients [Table/Fig-5].

In the present study we reported onychomycosis, an uncommon presentation of protothecosis. Among the rare presentations there are few cases of fungual protothecosis and as per the published reports this could be the first report from India which has been cultured proven and then confirmed by National Culture Collection by Pathogenic Fungi (NCCPF), PGI Chandigarh [21].

#### CONCLUSION

NCYG are emerging as important opportunistic fungal pathogens and are likely to be overlooked. They require high index of suspicion and clinical correlation. The study highlights the presence of these genera in clinical samples which may have therapeutic implications. Their significance and management would require case to case correlation by the clinical microbiologist and clinician and consequently clinicians need to have increased awareness of these fungal infections. Therefore, it is necessary to identify these genera. Automated identification systems for yeast-like genera could be of great use towards this end.

#### REFERENCES

- Gandham NR, Jadhav SV, Sardar M, Vyawahare CR, Misra RN. The spectrum and etiology of Mycotic infections from a tertiary care hospital from western part of India. JCDR. 2013;7(10):2157-59.
- [2] Chander J. Textbook of medical mycology. Third edition, Mehta publishers; 2012. Chapter 5 Diagnosis of fungal infections; Pp. 53-67.
- [3] Murray PR, Baron EJ, Jorgensen JH, Landry ML and Pfaller MA. Manual of Clinical Microbiology. 9<sup>th</sup> Edition. ASM Press, Washington DC. 2007 Chapter 119 Candida, Cryptococcus and other yeasts of medical importance 1762-1788.
- [4] Pradhan S, Agrawal E, Murthy R, Tomar R. Prevalence of invasive trichosporonosis by Trichosporonasahii and other Trichosporon species and their anti fungal susceptibility pattern in Chhattisgarh. Int J Med Res Rev. 2017;5(03):285-92.
- [5] Gandham NR, Vyawahare CR, Jadhav SV, Misra RN. Candidemia: Speciation and anti-fungal susceptibility testing from a tertiary care hospital in Maharashtra, India. Med. J DPU. 2016;9:596-99.
- [6] Verma S, Veerma GK, Singh G, Kanga A, Shanker V, Singh D, et al. Sporotrichosis in sub-himalayan India. PLoS Negl Trop Dis. 2012;6(6):e1673. https://doi. org.1371/journal.ptnd.0001673.
- [7] Guitard J, Angoulvant A, Letscher-Bru V, L'Oliver C, Cornet M, Dalle F, et al. Invasive infections due to Candida norvigensis and Candida inconspicua: report of 12 cases and review of literature. Med Mycol. 2013;51(8):795-99.
- [8] Wirth F, Goldani LZ. Epiodemiology of Rhodotorula: Emerging pathogen. Inter Persp Inf Dis. 2012;2012:465717.
- [9] Capoor MR, Gupta DK, Verma PK, Sachdeva HC. Rare yeasts causing fungemia in immunocompromised and hematology patients: case series from Delhi. Ind J Med Micro. 2015;33:576-79.
- [10] Ellis D, Davis S, Alexiou H, Handke R, Barley R. Description of Medical fungi, 2<sup>nd</sup> Ed. publishers authors; 2007. Chapter Trichosporon Behrend; p170-173.
- [11] Sugita T, Nakajima M, Ikeda R, Matsushima T, Shinoda T. Sequence analysis of the ribosomal DNA intergenic spacer 1 regions of Trichosporon species. J Clin Microbiol. 2002;40(5):1826-30.
- [12] Kalkanci A, Sugita T, Arikan S, Yucesoy M, Ener B, Otag F, et al. Molecular identification, genotyping and susceptibility of the Basidiomycetous yeast pathogen Trichosporon isolated from Turkish patients. Med Mycol. 2010;48(1):141-46.
- [13] Girmenia C, Pagano L, Martino B, D'Antonio D, Fanci R, Spechia G, et al. Invasive infections caused by Trichosporon species and Geotrichumcapitatum in patients with haematological malignancies: a retrospective multicenter study from Italy and review of literature. J Clin Microbiol. 2005;43(4):1818-28.
- [14] Vyawahare CR, Gandham NR, Misra RN, Jadhav SV, Gupta NS, Angadi KM. Occurence of catheter-associated urinary tract infection in critical care units. Med

Nageswari Rajesh Gandham et al., Non-candidial Yeast like Genera

J DPU. 2015;8(5):585-89.

- [15] Thomas C, Chagas N, Guilherme MC, Analy SAM, Amaldo LC. Blood stream infections due to Trichosporonspp: species distribution, Trichosporonasahi genotypes determined on the basis of Ribosomal DNA intergenic spacer! sequencing and antifungal susceptibility testing. J Clin Microbiol. 2009;47(4):1074-81.
- [16] Sood S, Pathak D, Sharma R, Rishi S. Urinary tract infection by Trichosporonasahii. Ind J Med Micro. 2006;24(4):294-96.
- [17] Khan ID, Sahni AK, Basu A, Haleem S. Trichosporonasahi urinary tract infection in immunocompetent patients. Med J Armed Forces Ind. 2015;71(4):373-76.
- [18] Shah A, Koticha A, Ubale M, Wanjare S, Mehta P, Khopkar U. Identification

and speciation of Malassezia in patients clinically suspected of having Pityriasis versicolor. Ind J Derm. 2013;58(3):239-44.

- [19] Kindo AJ, Sophia SK, Kalyani J, Anandan S. Identification of Malassezia species. Ind J Med Micro. 2004;22:179-81.
- [20] Batra R, Boekhout T, Gueho E, Cabanes J, Dawson TL, Gupta AK. Malasseziabaillon, emerging clinical yeasts. FEMS Yeast Research. 2005;5(12):1101-13.
- [21] Gandham NR, Vyawahare CR, Chaudhary N, Shinde RA. Onychoprotothecosis: An uncommon presentation of Protothecosis. Indian J Med Microbiol. 2015;33:435-37.

#### PARTICULARS OF CONTRIBUTORS:

- Professor, Department of Microbiology, Dr. D.Y. Patil Medical College, Hospital and Research Centre [Dr. D.Y. Patil Vidyapeeth Pune], Pimpri Pune, Maharashtra, India. Professor, Department of Microbiology, Dr. D.Y. Patil Medical College, Hospital and Research Centre [Dr. D.Y. Patil Vidyapeeth Pune], Pimpri Pune, Maharashtra, India. Professor, Department of Microbiology, Dr. D.Y. Patil Medical College, Hospital and Research Centre [Dr. D.Y. Patil Vidyapeeth Pune], Pimpri Pune, Maharashtra, India. 2. 3.
- Associate Professor, Department of Microbiology, Dr. D.Y. Patil Medical College, Hospital and Research Centre [Dr. D.Y. Patil Vidyapeeth Pune], Pimpri Pune, 4.
- Maharashtra India
- Associate Professor, Department of Microbiology, Dr. D.Y. Patil Medical College, Hospital and Research Centre [Dr. D.Y. Patil Vidyapeeth Pune], Pimpri Pune, 5. Maharashtra, India.

#### NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Dr. Savita Vivekanad Jadhav.

Professor, Department of Microbiology, Dr. D.Y. Patil Medical College, Hospital and Research Centre [Dr. D.Y. Patil Vidyapeeth Pune], Pimpri Pune, Maharashtra, India. E-mail: patilsv78@gmail.com

FINANCIAL OR OTHER COMPETING INTERESTS: None.

Date of Submission: Apr 12, 2018 Date of Peer Review: May 18, 2018 Date of Acceptance: Jun 08, 2018 Date of Publishing: Aug 01, 2018